UDC 616:619. 615.322-615.281.9:615.284-615.282.84-615.281.9

Original article

ANALYSIS OF ANTIPARASITIC ACTIVITY OF WILD PLANTS OF EASTERN AND NORTHERN KAZAKHSTAN

Kukhar Y.V.1*, Aidarkhanova G.S.2

- ¹Research Platform of Agricultural Biotechnology, S. Seifullin Kazakh Agrotechnical Research University, Astana, Kazakhstan ²Department of Biology, Plant Protection and Quarantine, S. Seifullin Kazakh Agrotechnical Research University, Astana, Kazakhstan
- * Corresponding Author: Kukhar Y.V., kucharev@mail.ru.

ABSTRACT

Various helminthoses and other parasitic infections are widespread among farm animals. Helminthic infections are typically treated with anthelmintic drugs; however, resistance to these drugs is becoming an increasingly serious problem. Investigating potential herbal antiparasitic agents, which are inexpensive and widely available, could be a promising alternative. Anthelmintic properties have been identified in oil, hydroalcoholic, and most aqueous extracts of *Filipendula vulgaris* Moench., *Pinus sylvestris*, *Picea abies* L., *Abies sibirica*, *Pinus sibirica* DuTour, *Juniperus communis* L., and *Paeonia anomala* L. from forest ecosystems in Eastern Kazakhstan, as well as in *Ferula tatarica* Fisch., *Pulsatilla uralensis* (Zămels) Tzvel., and *Serratula coronata* L. from ecosystems in Northern Kazakhstan. The greatest impact and fastest death of annelids were observed with alcohol tinctures, while aqueous extracts had the least effect. The best anthelmintic properties were found in the aqueous-alcoholic extracts of the underground part of *Pulsatilla uralensis* (Zămels) Tzvel. and the alcohol tincture of the aboveground part of *Serratula coronata* L. For the first time, the antiparasitic activity of both the aboveground and underground parts of *Pulsatilla uralensis* (Zămels) Tzvel., growing in Northern Kazakhstan, has been described

Keywords: antiparasitic activity, anthelmintic properties, coniferous trees, shrubs, herbaceous plants.

INTRODUCTION

Various helminthiases and other parasitic infections are widespread in farm animals [1]. Gastrointestinal parasitic diseases have become a major health issue, affecting billions of people and livestock worldwide, especially in developing countries. These diseases impact an estimated 3.5 billion people globally, with 450 million showing symptoms and over 200,000 deaths reported annually [2]. Gastrointestinal parasites in animals also create a significant disease burden and result in substantial economic losses in food production across many regions [3, 4].

Gastrointestinal helminths are a major cause of mortality and morbidity in ruminants, leading to high costs and production losses worldwide [5]. For instance, the annual cost of gastrointestinal nematode infections in Europe is estimated at £686 million [6]. One example of a gastrointestinal parasite affecting livestock is *Haemonchus contortus*, a highly virulent parasite of small ruminants in tropical, subtropical, and warm temperate regions [7, 8]. This parasite can cause ascites, weight loss, anemia, and death [9, 10, 11].

Protozoan parasites also pose a significant threat to the livestock industry, with *Cryptosporidium*, *Giardia*, and *Eimeria* species being among the most important. These infections affect a wide range of host animals worldwide, including cattle, sheep, goats, and pigs, leading to substantial growth and productivity losses [12]. Infected animals may exhibit clinical signs such as growth retardation, weight loss, decreased productivity, reduced appetite, dehydration, and diarrhea, which, in severe cases, can result in death [13, 14, 15, 16, 17, 18].

Helminthic infections are typically treated with anthelmintic drugs; however, resistance to these drugs is becoming an increasingly serious issue [1]. The growing evidence of multidrug resistance in these parasites, coupled with the adverse effects of currently available synthetic drugs, has spurred increased research into alternative treatments. The study of potential plant-based antiparasitic agents, which are inexpensive and widely available, presents a promising alternative [19].

Various plants, both medicinal and wild, are regularly analyzed for the presence of biologically active substances (BAS) with bactericidal, fungicidal, and helminthicidal properties. Coniferous plants, in particular, are well-known for their beneficial properties and effective BAS.

Coniferous trees contain a diverse range of biologically active substances in their bark, needles, and seeds. Among conifers, the Siberian fir is particularly noted for its medicinal properties and is a significant source of various phytochemicals with pharmacological importance. The pharmacological uses of fir include several effects, such as anthelmintic properties, especially when combined with other non-pharmacological applications [20]. Concentrated tannins in feed have been reported to reduce the helminth load in small ruminants. Bark extracts from pine (Pinus sylvestris L.), spruce (Picea abies L.), and birch (Betula pubescens) are good sources of tannins in cold climates [1]. Additionally, the seeds of Siberian pine or cedar pine (P. sibirica), commonly known as pine nuts, are valued for their medicinal properties due to the presence of both lipophilic and hydrophilic biologically active substances [21].

Shrubs, including coniferous ones, also offer valuable medicinal properties. For example, the needles of Siberian juniper (*Juniperus sibirica* L.) contain ADP, lignin, cellulose, lignin,

nin/cellulose, lipids, phenolic compounds, proanthocyanidins, flavonoids, and essential minerals such as N, C, Ca, Mg, K, Mn, Zn, P, S, Al, and Fe [22], as well as sabinene [23, 24], which have antioxidant, anti-inflammatory, and antimicrobial effects [25, 26].

Flowering plants also exhibit a wide range of biologically active substances. *Meadowsweet* (*Filipendula ulmaria*) is traditionally used for treating fever, pain, inflammatory diseases (such as arthrosis, rheumatism, and arthritis), gastric disorders, liver dysfunction, and gout. Phytochemical studies reveal several active compounds in *F. ulmaria*, including phenolic acids, flavonoids, tannins, terpenoids, and salicylic acid and its derivatives, which contribute to its diverse therapeutic activities [27].

Paeonia anomala L., a member of the Paeoniaceae family, is found in Asia, Europe, and western North America. Over 180 compounds have been isolated from nine species within the *Paeonia* genus, including terpenes, phenols, flavonoids, essential oils, and tannins. The rich phytochemical composition and bioactivity of peony, particularly due to its polyphenols and flavonoids, have been well-documented [28]. There are known differences in the content of biologically active substances between the above-ground and underground parts of plant shoots. Traditional healers and medicinal plant reference books often specify which part of the plant should be used to obtain a higher concentration of biologically active substances or the most effective components.

In Serratula coronata L., cultivated in Siberia, at least 14 phenolic compounds (7.3%) have been identified, including quercetin, which is highly effective in treating cerebral edema, gout, and cardiovascular diseases due to its angioprotective properties [29, 30]. The biocomponents of Serratula coronata L. have also proven effective in treating and preventing various skin diseases, particularly psoriasis [31, 32].

Several studies have explored the antiparasitic properties of *Ferula assa-foetida*, including its activity against *Trichomonas vaginalis* [33], *Schistosoma mansoni* [34], *Strongylus* spp. [35], *Fasciola hepatica*, and *Dicrocoelium dendriticum* [36]. Aqueous extracts of *Ferula foetida* resin were tested for anthelmintic activity against *Pheretima posthuma*, with three concentrations (25, 50, and 100 mg/ml) evaluated. The extract exhibited significant anthelmintic activity at 100 mg/ml, proving more effective than the drug Piperazine citrate [37].

The herb and roots of *Pulsatilla* species, or pasqueflower, are widely used for various therapeutic purposes. The extract from the leaves of *Pulsatilla pratensis* has strong bactericidal and fungicidal effects, provides sedation, and is used in both folk and veterinary medicine as a sedative, sleep aid, analgesic, expectorant, and liver function stimulant [38].

Studying the current state of forest plants in Kazakhstan is crucial for assessing their species diversity and resource potential. Given the increasing problem of parasite resistance to conventional anthelmintic drugs, developing herbal antiparasitic treatments from domestic raw materials is highly relevant. Therefore, investigating the anthelmintic effects of biologically active substances from wild forest plants in Kazakhstan seems particularly timely.

The aim of the research was to investigate the antiparasitic activity of wild medicinal plants, including both conifers and herbaceous perennials, to evaluate their pharmaceutical potential.

2. MATERIAL AND METHODS

he material for the study comprised 10 plant samples collected during the 2020 expedition. Samples of Scots pine (*Pinus sylvestris*), Norway spruce (*Picea abies* L.), Siberian fir (*Abies sibirica*), Siberian cedar pine (*Pinus sibirica* DuTour), common juniper (*Juniperus communis* L.), peony (*Paeonia anomala* L.), and meadowsweet (*Filipendula vulgaris* Moench) were collected from the forest ecosystems of the mountainous region in the Kazakhstan part of Altai. Additionally, samples of *Ferula tatarica* Fisch., *Pulsatilla uralensis* (Zămels) Tzvel., and *Serratula coronata* L. were gathered from forested areas in Northern Kazakhstan.

During sample collection, the coordinates of each site were recorded, geobotanical descriptions were made, and species were identified using established plant identification guides [39, 40].

The biological activity of the extracts was analyzed at the Research Platform for Agricultural Biotechnology of the S. Seifullin Kazakh Agrotechnical Research University. The plant samples were dried to an air-dry state on shelves indoors. Averaged samples were prepared by grinding both the above-ground and underground parts and thoroughly mixing them. From these averaged plant samples, oil, alcohol, and water extracts were prepared following the State Pharmacopoeia of the Republic of Kazakhstan (2008) [41].

Infusions and tinctures were allowed to infuse for 2 weeks in a dark place, while decoctions were prepared immediately before use. Sterilizing filtration of the preparations was performed using filters with a pore diameter of 0.45 μ m. The extracts were stored at 2-8°C in a household refrigerator for no more than three days.

The anthelmintic activity of the plant materials was assessed using a modified technique on annelids – specifically, ringed worms (*Lumbricus terrestris*), which served as the test objects. Observations of worm behavior in Petri dishes containing solid nutrient medium with wells filled with the extracts were conducted at 3, 6, and 12 hours on the first day, and every 8 hours over the next two days. A corresponding extractant served as the negative control, and a solution of piperazine citrate (20 mg/100 ml) was used as the positive control.

The presence of anthelmintic effects was evaluated based on the following criteria: natural behavior of the worms, their tendency to approach or move away from the wells, death within a certain period, the presence and intensity of decomposition odor if death occurred, and the presence and intensity of hemolysis. Each criterion was recorded with the following notation: +++ for pronounced effect, ++ for moderate effect, + for slight effect, and – for no effect. Based on the results, the total number of "+" signs were calculated, and points were assigned [42]. The results were interpreted using standard statistical methods.

3. RESULTS

The results of the study on the antiparasitic properties of plant extracts from coniferous trees are presented in Figure 1.

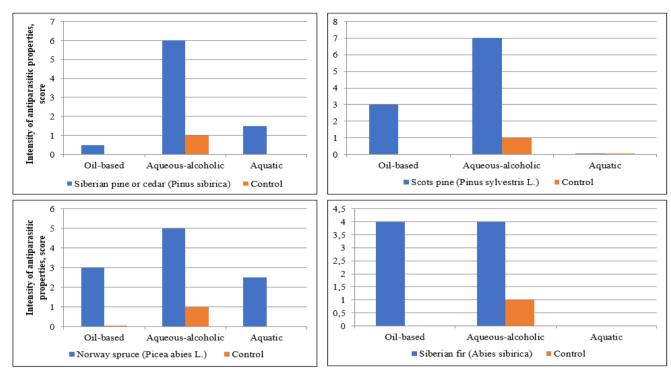


Figure 1 – Intensity of anthelmintic properties of various coniferous tree extracts.

The data indicate that plant extracts from wild coniferous forest trees exhibit antiparasitic properties. Among the extracts, alcohol tinctures from Scots pine and Siberian pine (cedar pine) demonstrated the most significant antiparasitic effects. Conversely, Siberian fir showed the least pronounced antiparasitic properties. Of the oil extracts, Siberian fir was the most effective against *Lumbricus terrestris*. Norway spruce and Scots pine extracts exhibited moderate effectiveness, while cedar pine had minimal antiparasitic impact.

Among the aqueous solutions, Scots spruce and cedar pine extracts showed a weak effect on worms, while aqueous extracts of Scots pine and Siberian fir had no impact on the behavior of *Lumbricus terrestris*. Additionally, extracts of various chemical natures obtained from coniferous shrubs, including juniper, also displayed varying degrees of antiparasitic activity (Figure 2).

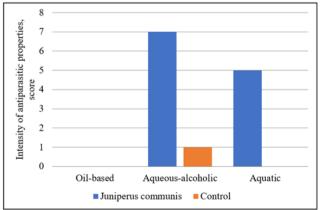


Figure 2 – Intensity of anthelmintic properties of various extracts of *Juniperus communis* L. (common juniper).

The results for plant extracts of common juniper show that alcohol tinctures and water extracts were the most effective, while the oil extract of common juniper had no effect on the behavior of *Lumbricus terrestris*.

The antiparasitic properties of herbaceous plant extracts from *Paeonia anomala* L. (peony) and common meadow-sweet (*Filipendula vulgaris*) are presented in Figure 3.

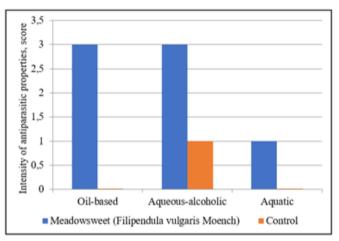
The data on the antiparasitic properties of peony extracts indicate that alcohol tinctures produced the best results. These tinctures caused convulsions, a tendency for worms to move away from the wells, and rapid death. Hemolysis and rapid decomposition were observed post-mortem. Peony oil extracts also showed a marked effect, causing hemolysis and rapid decomposition. In contrast, water infusions of peony did not affect earthworms.

Meadowsweet preparations, despite their relatively weak anthelmintic effect, demonstrated specific activity, leading to worm mortality by the end of the third day. Both oil and alcohol extracts of meadowsweet exhibited similar levels of anthelmintic activity, while the water extract was twice as effective as the water infusion of peony.

The anthelmintic properties of extracts from the above-ground and underground parts of *Pulsatilla uralensis* (Zămels) Tzvel. (yellow pasqueflower), *Ferula tatarica* Fisch. ex Spreng (Tatar ferula), and *Serratula coronata* L. (crowned sickleweed) on *Lumbricus terrestris* are presented in Figure 4.

As shown in the diagram, the best anthelmintic properties were demonstrated by the extracts from the underground parts of the analyzed plants. Among these, the aqueous-alcoholic extracts of both the above-ground and underground parts of *Pulsatilla uralensis* (Zămels) Tzvel. (yellow pasqueflower) were more effective than the oil and water extracts. Notably, the alcoholic tincture of the above-ground part of *Serratula coronata* L. (crowned sickleweed), along with the aqueous-alcoholic extract of the underground part of *Pulsatilla uralensis*, exhibited some of the highest antiparasitic effects, leading to very rapid death of the worms.

The intensity of these antiparasitic properties was especially pronounced, even in the presence of a positive control



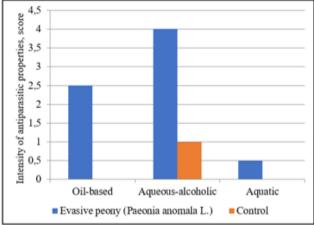
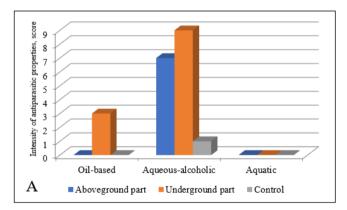


Figure 3 – Intensity of anthelmintic properties of various extracts of *Paeonia anomala* L. (peony) and *Filipendula vulgaris* Moench. (meadowsweet).

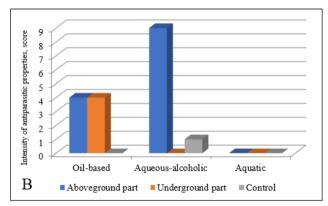
(an aqueous solution of ethyl alcohol). Oil extracts of *Serratula coronata* also demonstrated a significant antiparasitic effect, with worms dying within 6 hours of initial observation. In contrast, aqueous extracts from both the above-ground and underground parts of *Serratula coronata* showed no antiparasitic activity.

All extracts of *Ferula tatarica* Fisch. ex Spreng exhibited minimal impact on the behavior of *Lumbricus terrestris*, although they did cause worm mortality. The most pronounced effect and rapid death of annelids were achieved with alcohol tinctures, while oil extracts were less effective.



4. DISCUSSION

From the analysis of available literature, it is evident that several plants exhibit antiparasitic and anthelmintic properties. For example, extracts from the bark of various tree species, including pine (*Pinus sylvestris* L.), spruce (*Picea abies* L.), and birch (*Betula pubescens*), have been tested for their anthelmintic efficacy against the infectious nematode of sheep, *Teladorsagia circumcincta*. These studies demonstrated the anthelmintic activity of both aqueous and acetone extracts of these tree barks through two independent in vitro assays: egg hatching and larval motility assays [1].



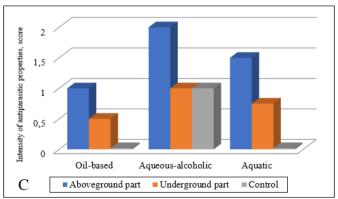


Figure 4 – Intensity of anthelmintic properties of various extracts from the above-ground and underground parts of herbaceous plants: A - Yellow pasqueflower (*Pulsatilla uralensis* (Zămels) Tzvel.), B - Crowned sickleweed (*Serratula coronata* L.), C - Tatarian ferula (*Ferula tatarica* Fisch. ex Spreng).

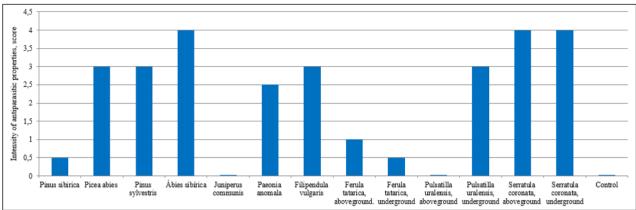


Figure 5 – Comparative analysis of the anthelmintic properties of oil extracts of the studied plants.

Siberian fir is noted for its anthelmintic properties, among other effects, in combination with other non-pharmacological uses [20]. Siberian juniper (*Juniperus sibirica* L.), when used as a decoction, is recommended for rheumatism and arthritis, while an infusion is suggested for neuroses, liver, and kidney diseases [21]. Essential oils from Siberian juniper needles have demonstrated antibacterial properties and a bacteriostatic effect against certain gram-negative bacteria [43], though an anthelmintic effect has not been described.

Pharmacological assessments of *Filipendula ulmaria* (meadowsweet) have highlighted its anticoagulant, antiarthritic, analgesic, anti-inflammatory, antioxidant, anticancer, antimicrobial, immunomodulatory, gastroprotective, and hepatoprotective activities [29]. The presence of anthelmintic action has been noted for phenolic compounds from *Filipendula ulmaria*, as well as from *Agrimonia spp*. (burdock) and *Crataegus spp*. (hawthorn) [44].

assays and scanning electron microscopy (SEM). SEM images of *F. assa-foetida* extract (200 μg/ml) treated worms showed excessive damage, including the complete loss of sensory papillae and destruction of discernible network structures and tegument vesicles. Increased time and concentration led to higher mortality rates among trematodes, confirming the anthelmintic properties of *F. assa-foetida* [36].

Pulsatilla pratensis is known for its anti-inflammatory, antibacterial, antifungal, antimicrobial, antimalarial, and antitumor properties. However, data on the composition and biological properties of Pulsatilla uralensis growing in Kazakhstan are limited and focus mainly on the antimicrobial properties of its essential oils [38, 47]. Our analysis shows that almost all studied oil extracts of wild plants negatively affect the vitality of annelids, specifically Lumbricus terrestris, used as the test object (Figure 5).

According to Figure 5, nearly all oil extracts from the

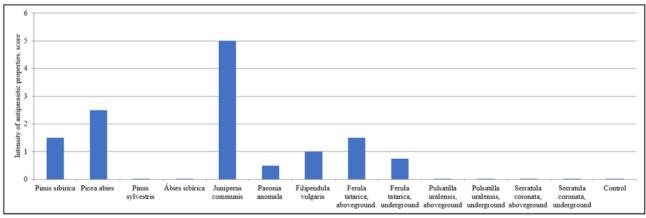


Figure 6 – Comparative analysis of the anthelmintic properties of aqueous extracts of the studied plants.

Paeonia anomala L. exhibits a range of pharmacological activities, including antibacterial, antifungal, anticoagulant, inhibitory, phytotoxic, and insecticidal effects, along with lipoxygenase and beta-glucuronidase activities, and radical scavenging activity [45]. However, the anthelmintic effects of Kazakhstani populations of Paeonia anomala have not been described.

The active components of *Serratula coronata* L. extracts are known to exhibit various biological effects, including antiparasitic properties [46]. The anthelmintic effects of *Ferula assa-foetida* extracts against *Fasciola hepatica* and *Dicrocoelium dendriticum* were confirmed through in vitro

studied plants demonstrate significant antiparasitic activity. Exceptions include the oil extracts of Siberian pine (*Pinus sibirica*), Tatar ferula (*Ferula tatarica*, underground part), and yellow pasqueflower (*Pulsatilla uralensis*, aerial part), which show minimal antiparasitic effects. In comparison, the water extracts of the studied plants exhibit a less pronounced antiparasitic effect on the behavior of *Lumbricus terrestris* (Figure 6).

As shown in Figure 6, the aqueous extract of common juniper demonstrated notable antiparasitic activity. In contrast, the aqueous extracts of Siberian pine, common spruce, peony, meadowsweet, and both aboveground and under-

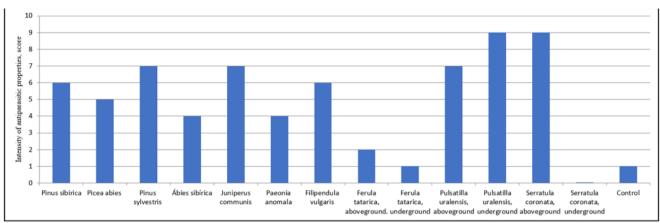


Figure 7 – Comparative analysis of the anthelmintic properties of aqueous-alcoholic extracts of the studied plants.

ground parts of *Ferula tatarica* exhibited only weak activity. The remaining aqueous extracts showed no antiparasitic activity.

Among all the wild plant preparations studied, aqueous-alcoholic extracts exhibited the best antiparasitic activity. Specifically, the aqueous-alcoholic extracts from the aboveground vegetative organs of *Serratula coronata L*. and the underground vegetative organs of *Pulsatilla uralensis* (Zămels) Tzvel. showed the most pronounced antiparasitic effects. These extracts caused convulsions, paralysis, and nearly instantaneous death of *Lumbricus terrestris* in Petri dishes. The antiparasitic effects of extracts from *Pinus sylvestris*, *Juniperus communis*, and the aerial parts of *Pulsatilla uralensis* were somewhat weaker (Figure 7).

Figure 7 shows that alcoholic extracts of *Ferula tatarica* and the underground parts of *Serratula coronata* exhibited minimal antiparasitic activity. The remaining aqueous-alcoholic tinctures produced average results, with their effects manifesting in the death of *Lumbricus terrestris* by the second day.

These findings are inconsistent with reports by Ramadan N.I. et al. (2004), Tavassoli M. et al. (2018), Gundamaraju R. (2013), and others, who identified pronounced anthelmintic properties in various species of *Ferula* (e.g., *Ferula asafetida*, *Ferula foetida*) [33-37]. This discrepancy may be attributed to regional climatic conditions or other factors, warranting further investigation.

From the data, it is evident that the most effective antiparasitic activity was observed in alcohol tinctures from the following plants: *Pulsatilla uralensis* (both aerial parts and roots), *Serratula coronata* (aerial parts), *Pinus sylvestris*, and *Juniperus communis*. Among aqueous solutions, *Juniperus communis* demonstrated a notable antiparasitic effect, whereas *Picea abies* exhibited weaker activity. In oil extracts, *Ábies sibírica* and *Serratula coronata* (both aerial and underground parts) were the most effective.

These findings align with previous studies indicating anthelmintic and antiparasitic properties in similar plant extracts [1, 34, 37-41]. Plants collected from the Altai Botanical Garden (Ridder) and the mountain forests of Western Altai in Eastern Kazakhstan displayed anthelmintic activity, though not as pronounced as those from Northern Kazakhstan.

Particularly noteworthy is the antiparasitic activity of *Pulsatilla uralensis* against *Lumbricus terrestris*, which is re-

ported here for the first time. The pronounced biological activity observed in these plant extracts suggests their potential for use in developing domestic plant-based remedies for parasitic diseases.

CONCLUSION

Anthelmintic Properties: Anthelmintic properties were identified in oil, water-alcohol, and most water extracts of Filipendula vulgaris Moench., Pinus sylvestris, Picea abies L., Ábies sibírica, Pinus sibirica DuTour, Juniperus communis L., and Paeonia anomala L. from forest ecosystems of Eastern Kazakhstan. Additionally, Ferula tatarica Fisch., Pulsatilla uralensis (Zămels) Tzvel., and Serratula coronata L. from Northern Kazakhstan ecosystems also exhibited anthelmintic properties. Alcohol tinctures demonstrated the greatest impact, causing rapid death in annelids, whereas water extracts were the least effective.

Most Effective Extracts: The most effective anthelmintic properties were observed in aqueous-alcoholic extracts of the underground parts of *Pulsatilla uralensis* (Zămels) Tzvel., and alcohol tinctures of the aboveground parts of *Serratula coronata* L., which exhibited some of the highest levels of antiparasitic activity.

Novel Findings: The antiparasitic activity of both the aboveground and underground parts of *Pulsatilla uralensis* (Zămels) Tzvel., from Northern Kazakhstan, is reported here for the first time.

ACKNOWLEDGMENTS

The authors express their gratitude to Nysanbek Zh. for conducting the laboratory tests essential to this study.

FUNDING

This study was carried out within the framework of Project AP05136154 "Resource Potential of Non-Wood Forest Materials and Their Environmental Safety for the Socio-Economic Development of the Regions of Kazakhstan," financed by the Ministry of Education and Science of the Republic of Kazakhstan during 2018-2020.

CONFLICT OF INTEREST

There are no conflicts of interest to declare.

LITERATURE

- 1. Athanasiadou, S., Almvik, M., Hellström, J., Madland, E., Simic, N., Steinshamn, H. Chemical Analysis and Anthelmintic Activity Against Teladorsagia Circumcincta of Nordic Bark Extracts In vitro // Frontiers in veterinary science. 2021. Vol. 8. P. 666924. https://doi.org/10.3389/fvets.2021.666924.
- 2. Essential Nutrition Actions: Improving Maternal, Newborn, Infant and Young Child Health and Nutrition. (2013). World Health Organization. 2016. Retrieved from https://apps.who.int/iris/bitstream/handle/10665/326261/9789241515856-eng.pdf?ua=1.
- 3. Mehlhorn, H. Encyclopedia of parasitology: volume 1, AM. 3Edn ed. Heidelberg, Germany: Springer Science & Business Media, 2008.
- 4. Roeber, F., Jex, A.R., & Gasser, R.B. Impact of gastrointestinal parasitic nematodes of sheep, and the role of advanced molecular tools for exploring epidemiology and drug resistance an Australian perspective // Parasites & vectors. 2013. Vol. 6. P. 153. https://doi.org/10.1186/1756-3305-6-153.
- 5. Perry, B.D., Ranolph, T.F., McDermott, J.J., Sones, K.R., Thornton P.K. Investing in animal health research to alleviate poverty. International Livestock Research Institute, Nairobi, Kenya. 2002. P. 17.
- 6. Charlier, J., Rinaldi, L., Musella, V., Ploeger, H.W., Chartier, C., Vineer, H.R., Hinney, B., von Samson-Himmelstjerna, G., Băcescu, B., Mickiewicz, M., Mateus, T.L., Martinez-Valladares, M., Quealy, S., Azaizeh, H., Sekovska, B., Akkari, H., Petkevicius, S., Hektoen, L., Höglund, J., Morgan, E.R., Claerebout, E. Initial assessment of the economic burden of major parasitic helminth infections to the ruminant livestock industry in Europe // Preventive veterinary medicine. 2020. Vol. 182. P. 105103. https://doi.org/10.1016/j.prevetmed.2020.105103.
- 7. Qamar, M.F., Maqbool, A., Khan, M.S., Ahmad, N., Muneer, M.A. Epidemiology of haemonchosis in sheep and goats under different managemental conditions // Vet World. -2009. Vol. 2(11). P. 413-417.
- 8. Sissay, M.M., Uggla, A., Waller, P.J. Prevalence and seasonal incidence of nematode parasites and fluke infections of sheep and goats in eastern Ethiopia // Tropical animal health and production. 2007. Vol. 39(7). P. 521-531. https://doi.org/10.1007/s11250-007-9035-z.
- 9. Roeber, F., Jex, A.R., Gasser, R.B. Impact of gastro-intestinal parasitic nematodes of sheep, and the role of advanced molecular tools for exploring epidemiology and drug resistance an Australian perspective // Parasites & vectors. 2013. Vol. 6. P. 153. https://doi.org/10.1186/1756-3305-6-153.
- 10. Abakar, A, Osman, A. Haematological and biochemical changes following concurrent infections with coccidia and *Haemonchus contortus* in desert lambs // J. Anim. Vet. Adv. 2004. Vol. 3(10). P. 643-647.
- 11. Kelkele, F.A., Tolossa, Y.H., Kassa, G.M. Experimental infection of Ethiopian highland sheep by different infective doses of *Haemonchus contortus* (L3): haematological and parasitological parameters, serum protein con-

- centrations and clinical responses // Ethiop. Vet. J. 2012. Vol. 6(1). P. 41-57. https://doi.org/10.4314/evj.v16i1.4.
- 12. Robertson, L.J., Björkman, C., Axén, C., Fayr, R. Cryptosporidiosis in farmed animals. In book: Cryptosporidium: parasite and disease. Vienna: Springer, 2014. P. 149-235.
- 13. Thomson, S., Hamilton, C.A., Hope, J.C., Katzer, F., Mabbott, N.A., Morrison, L.J., Innes, E.A. Bovine cryptosporidiosis: impact, host-parasite interaction and control strategies // Veterinary research. 2017. Vol. 48(1). P. 42. https://doi.org/10.1186/s13567-017-0447-0.
- 14. Klein, P., Kleinová, T., Volek, Z., Simůnek, J. Effect of Cryptosporidium parvum infection on the absorptive capacity and paracellular permeability of the small intestine in neonatal calves // Veterinary parasitology. 2008. Vol. 152 (1-2). P. 53-59. https://doi.org/10.1016/j.vet-par.2007.11.020.
- 15. Feng, Y., Xiao, L. Zoonotic potential and molecular epidemiology of Giardia species and giardiasis // Clinical microbiology reviews. 2011. Vol. 24(1). P. 110-140. https://doi.org/10.1128/CMR.00033-10.
- 16. Santin, M. Cryptosporidium and Giardia in Ruminants. The Veterinary clinics of North America // Food animal practice. 2020. Vol. 36(1). P. 223-238. https://doi.org/10.1016/j.cvfa.2019.11.005.
- 17. Cacciò, S.M., Lalle, M., Svärd, S.G. Host specificity in the *Giardia duodenalis* species complex. Infection, genetics and evolution // Journal of molecular epidemiology and evolutionary genetics in infectious diseases. 2018. Vol. 66. P. 335-345. https://doi.org/10.1016/j.mee-gid.2017.12.001.
- 18. Daugschies, A., Najdrowski, M. Eimeriosis in cattle: current understanding // Journal of veterinary medicine. B, Infectious diseases and veterinary public health. 2005. Vol. 52(10). P. 417-427. https://doi.org/10.1111/j.1439-0450.2005.00894.x.
- 19. Ranasinghe, S., Armson, A., Lymbery, A.J., Zahedi, A., Ash, A. Medicinal plants as a source of antiparasitics: an overview of experimental studies // Pathogens and global health. 2023. Vol. 117(6). P. 535-553. https://doi.org/10.1080/20477724.2023.2179454.
- 20. Assad, R., Reshi, Z.A., Mir, S.H., Rashid, I., Shouche, Y., Dhotre, D. Chapter 17 Bioprospecting appraisal of Himalayan pindrow fir for pharmacological applications, Academic Press, 2021. P. 461-482. https://doi.org/10.1016/B978-0-12-824109-7.00003-0.
- 21. Lantto, T.A., Dorman, H.J.D., Shikov, A.N., Pozharitskaya, O.N., Makarov, V.G., Tikhonov, V.P., Hiltunen, R., Raasmaja, A. Chemical composition, antioxidative activity and cell viability effects of a Siberian pine (*Pinus sibirica* Du Tour) extract // Food Chemistry. 2009. Vol. 112 (4). P. 936-943. https://doi.org/10.1016/j.foodchem.2008.07.008.
- 22 Artemkina, N.A., Orlova, M.A. Lukina, N.V. Chemical composition of *Juniperus sibirica* needles (*Cupressaceae*) in the forest-tundra ecotone, the Khibiny Mountains // Russ. J. Ecol. 2016. Vol. 47. P. 321-328. https://doi.org/10.1134/S106741361604007X.

- 23. Sampietro, D.A., Gomez, A.L., Jimenez, C.M., Lizarraga, E.F., Ibatayev, Z.A., Suleimen, Y.M., Catalán, C.A. Chemical composition and antifungal activity of essential oils from medicinal plants of Kazakhstan // Natural product research. - 2017. -Vol. 31(12). - P. 1464-1467. https://doi.org/ 10.1080/14786419.2016.1258560.
- 24. Caramiello, R., Bocco, A., Buffa G., Maffei M. Chemotaxonomy of Juniperus communis, J. sibirica and J. intermedia // Journal of Essential Oil Research. - 1995. - Vol. 7(2). – P. 133-145. https://doi.org/10.1080/10412905.1995.9 698488.
- 25. Lesjaka, M.M., Bearaa, I.N., Orčića, D.Z., Anačkovb, G.T., Baloga, K.J., Franciškovića, M.M., Mimica-Dukića, N.M. Juniperus sibirica Burgsdorf. as a novel source of antioxidant and anti-inflammatory agents // Food Chemistry. – 2011. – Vol. 124 (3). – P. 850-856. https://doi.org/10.1016/j. foodchem.2010.07.006.
- 26. Zhang, Y., Wu, D., Kuang, S., Qing, M., Ma, Y., Yang, T., Wang, T., Li, D. Chemical composition, antioxidant, antibacterial and cholinesterase inhibitory activities of three Juniperus species // Natural product research. – 2020. – Vol. 34(24). - P. 3531-3535. https://doi.org/10.1080/14786419.2 019.1579811.
- 27. Farzaneh, A., Hadjiakhoondi, A., Khanavi, M., Manayi, A., Bahramsoltani, R., Kalkhorani M. Filipendula ulmaria (L.) Maxim. (Meadowsweet): a Review of Traditional // Research Journal of Pharmacognosy (RJP). - 2022. - Vol. 9(3). - P. 85-106. https://doi.org/10.22127/ RJP.2021.302028.1781.
- 28. <u>Čutović N., Marković T., Kostić M., roš Gašić U., Pri-</u> jić Ž., Ren X., Lukić M., Bugarski B. Chemical Profile and Skin-Beneficial Activities of the Petal Extracts of Paeonia tenuifolia L. from Serbia // Pharmaceuticals (Basel). - 2022. -Vol. 15(12). – P. 1537. https://doi.org/10.3390/ph15121537.
- 29. Day, A.J., Bao, Y., Morgan, M.R., Williamson, G. Conjugation position of quercetin glucuronides and effect on biological activity // Free radical biology & medicine. – 2000. - Vol. 29(12). - P. 1234-1243. https://doi.org/10.1016/s0891-5849(00)00416-0.
- 30. Lupanova I.A, Saybel O.L., Ferubko E.V., Mizina P.G. Pharmacological screening of Serratula coronata L. herbal extract // Problems of Biological Medical and Pharmaceutical <u>Chemistry</u> January. – 2022. – Vol. 25(3). – P. 43-48. https:// doi.org/10.29296/25877313-2022-03-06.
- 31. Kroma, A., Pawlaczyk, M., Feliczak-Guzik, A., Urbańska, M., Jenerowicz, D., Seraszek-Jaros, A., Kikowska, M., Gornowicz-Porowska, J. Phytoecdysteroids from Serratula coronata L. for Psoriatic Skincare // Molecules (Basel, Switzerland). – 2022. – Vol. 27(11). – P. 3471. https://doi. org/10.3390/molecules27113471.
- 32. Odinokov, V.N., Galyautdinov, I.V., Nedopekin, D.V., Khalilov, L.M., Shashkov, A.S., Kachala, V.V., Dinan, L., Lafont, R. Phytoecdysteroids from the juice of Serratula coronata L. (Asteraceae) // Insect biochemistry and molecular biology. – 2002. – Vol. 32(2). – P. 161-165. https://doi. org/10.1016/s0965-1748(01)00106-0.
- 33. Vazini, H, Rahimi Esboei, B. In vitro study of the effect of hydroalcholic extracts of Carum copticum and Ferula asafetida against Trichomonas vaginalis // Scientific Journal

- of Kurdistan University of Medical Sciences. 2018. Vol. 23(1). – P. 52-61. https://doi.org/10.52547/sjku.23.1.76.
- 34. Ramadan, N.I., Abdel-Aaty, H.E., Abdel-Hameed, D.M., El Deeb, H.K., Samir, N.A., Mansy, S.S., Al Khadrawy, F.M. Effect of Ferula assafoetida on experimental murine Schistosoma mansoni infection // Journal of the Egyptian Society of Parasitology. - 2004. - Vol. 34(3 Suppl). - P. 1077-1094.
- 35. Tavassoli, M., Jalilzadeh-Amin, G., Fard, V. R. B., Esfandiarpour, R. The in vitro effect of Ferula asafoetida and Allium sativum extracts on Strongylus spp // Annals of parasitology. – 2018. – Vol. 64(1). – P. 59-63. https://doi.org/10.17420/ ap6401.133.
- 36. Arbabi, M., Haddad, A., Esmaeli, M., Hooshyar, H., Sehat, M. In Vitro Anthelmintic Effect of Ferula assa-foetida Hydroalcoholic Extract Against Flukes of Fasciola hepatica and Dicrocoelium dendriticum // Jundishapur J. Nat. Pharm. Prod. – 2023. – Vol. 18(1). – P. e133030. https://doi. org/10.5812/jjnpp-133030.
- 37. Gundamaraju, R. Evaluation of anti-helmintic activity of Ferula foetida "Hing-A natural Indian spice" aqueous extract // Asian Pac. J. Trop. Dis. – 2013. – Vol. 3(3). – P. 189-91. https://doi.org/10.1016/S2222-1808(13)60038-9
- 38. Kirillov, V.Yu., Stikhareva, T.N., Serafimovich, M.V., Mukasheva, F.T., Gering, A.V., Sarsenbaeva, L.A., Atazhanova, G.A., Adekenov, S.M. Chemical composition of essential oil from two species of Pulsatilla growing wild in Northern Kazakhstan // Bulletin of the Karaganda university. − 2018. − Vol. 2(90). − P. 29-34.
- Байтенов М.С. Флора Казахстана. Иллюстрированный определитель семейств и родов. -Алматы: Алматы кiтап, 1978. – 305 с.
- 40. The Plant List. Version 1.1. Electronic resource: http:// www.theplantlist.org.
- 41. State Pharmacopeia of Kazakhstan Republic. Almaty, 2008. – Vol. 1. – P. 361-401.
- 42. Aidarkhanova, G.S. Research report (AP05136154), Nur-Sultan, 2021. – 109 pp. (in Russian).
- 43. Nikolić, B., Vasiljević, B., Mitić-Ćulafić, D., Lesjak, M., Vuković-Gačić, B., Mimica Dukić, N., Knežević-Vukčević, J. Screening of the antibacterial effect of Juniperus sibirica and Juniperus sabina essential oils in a microtitre platebased MIC assay // Botanica Serbica. – 2016. – Vol. 40(1). - P. 43-48. https://doi.org/10.5281/zenodo.48858.
- 44. Elaine M Aldred BSc (Hons), DC, Lic Ac, Dip Herb Med, Dip CHM. Pharmacology: A Handbook for Complementary Healthcare Professionals, 2009. https://doi.org/10.1016/ B978-0-443-06898-0.X0001-1.
- 45. Zahra, N., Iqbal, J., Arif, M., Abbasi, B.A., Sher, H., Nawaz, A.F., Yaseen, T., Ydyrys, A., Sharifi-Rad, J., Calina, D. A comprehensive review on traditional uses, phytochemistry and pharmacological properties of Paeonia emodi Wall. ex Royle: current landscape and future perspectives // Chinese medicine. – 2023. – Vol. 18(1). – P. 23. https://doi. org/10.1186/s13020-023-00727-7.
- 46. Akhmetkarimova, Z., Temirgaziev, B. Phytoecdysteroids: Chemical Structure And Biological Activity // Eurasian Journal of Applied Biotechnology. - 2021. - Vol. 2. - P.

3-22. https://doi.org/10.11134/btp.2.2021.1.

47. Aidarkhanova, G., Kubentayev, S.A., <u>Kukhar, Ye.V.</u>, Urazalina, A.S. Current state of populations, resources, and biological activity *Pulsatilla patens* (Ranunculaceae) in Northern Kazakhstan // Biodiversitas. – 2022. – Vol. 23 (5). – P. 2311-2320. https://doi.org/10.13057/biodiv/d230509.

REFERENCES

- 1. Athanasiadou, S., Almvik, M., Hellström, J., Madland, E., Simic, N., Steinshamn, H. Chemical Analysis and Anthelmintic Activity Against Teladorsagia Circumcincta of Nordic Bark Extracts In vitro // Frontiers in veterinary science. 2021. Vol. 8. P. 666924. https://doi.org/10.3389/fvets.2021.666924.
- 2. Essential Nutrition Actions: Improving Maternal, Newborn, Infant and Young Child Health and Nutrition. (2013). World Health Organization. 2016. Retrieved from https://apps.who.int/iris/bitstream/handle/10665/326261/9789241515856-eng.pdf?ua=1.
- 3. Mehlhorn, H. Encyclopedia of parasitology: volume 1, AM. 3Edn ed. Heidelberg, Germany: Springer Science & Business Media, 2008.
- 4. Roeber, F., Jex, A.R., & Gasser, R.B. Impact of gastrointestinal parasitic nematodes of sheep, and the role of advanced molecular tools for exploring epidemiology and drug resistance an Australian perspective // Parasites & vectors. 2013. Vol. 6. P. 153. https://doi.org/10.1186/1756-3305-6-153.
- 5. Perry, B.D., Ranolph, T.F., McDermott, J.J., Sones, K.R., Thornton P.K. Investing in animal health research to alleviate poverty. International Livestock Research Institute, Nairobi, Kenya. 2002. P. 17.
- 6. Charlier, J., Rinaldi, L., Musella, V., Ploeger, H.W., Chartier, C., Vineer, H.R., Hinney, B., von Samson-Himmelstjerna, G., Băcescu, B., Mickiewicz, M., Mateus, T.L., Martinez-Valladares, M., Quealy, S., Azaizeh, H., Sekovska, B., Akkari, H., Petkevicius, S., Hektoen, L., Höglund, J., Morgan, E.R., Claerebout, E. Initial assessment of the economic burden of major parasitic helminth infections to the ruminant livestock industry in Europe // Preventive veterinary medicine. 2020. Vol. 182. P. 105103. https://doi.org/10.1016/j.prevetmed.2020.105103.
- 7. Qamar, M.F., Maqbool, A., Khan, M.S., Ahmad, N., Muneer, M.A. Epidemiology of haemonchosis in sheep and goats under different managemental conditions // Vet World. -2009. Vol. 2(11). P. 413-417.
- 8. Sissay, M.M., Uggla, A., Waller, P.J. Prevalence and seasonal incidence of nematode parasites and fluke infections of sheep and goats in eastern Ethiopia // Tropical animal health and production. 2007. Vol. 39(7). P. 521-531. https://doi.org/10.1007/s11250-007-9035-z.
- 9. Roeber, F., Jex, A.R., Gasser, R.B. Impact of gastro-intestinal parasitic nematodes of sheep, and the role of advanced molecular tools for exploring epidemiology and drug resistance an Australian perspective // Parasites & vectors. 2013. Vol. 6. P. 153. https://doi.org/10.1186/1756-3305-6-153.
 - 10. Abakar, A, Osman, A. Haematological and biochem-

- ical changes following concurrent infections with coccidia and *Haemonchus contortus* in desert lambs // J. Anim. Vet. Adv. 2004. Vol. 3(10). P. 643-647.
- 11. Kelkele, F.A., Tolossa, Y.H., Kassa, G.M. Experimental infection of Ethiopian highland sheep by different infective doses of *Haemonchus contortus* (L3): haematological and parasitological parameters, serum protein concentrations and clinical responses // Ethiop. Vet. J. 2012. Vol. 6(1). P. 41-57. https://doi.org/10.4314/evj.v16i1.4.
- 12. Robertson, L.J., Björkman, C., Axén, C., Fayr, R. Cryptosporidiosis in farmed animals. In book: Cryptosporidium: parasite and disease. Vienna: Springer, 2014. P. 149-235.
- 13. Thomson, S., Hamilton, C.A., Hope, J.C., Katzer, F., Mabbott, N.A., Morrison, L.J., Innes, E.A. Bovine cryptosporidiosis: impact, host-parasite interaction and control strategies // Veterinary research. 2017. Vol. 48(1). P. 42. https://doi.org/10.1186/s13567-017-0447-0.
- 14. Klein, P., Kleinová, T., Volek, Z., Simůnek, J. Effect of Cryptosporidium parvum infection on the absorptive capacity and paracellular permeability of the small intestine in neonatal calves // Veterinary parasitology. 2008. Vol. 152 (1-2). P. 53-59. https://doi.org/10.1016/j.vet-par.2007.11.020.
- 15. Feng, Y., Xiao, L. Zoonotic potential and molecular epidemiology of Giardia species and giardiasis // Clinical microbiology reviews. 2011. Vol. 24(1). P. 110-140. https://doi.org/10.1128/CMR.00033-10.
- 16. Santin, M. Cryptosporidium and Giardia in Ruminants. The Veterinary clinics of North America // Food animal practice. 2020. Vol. 36(1). P. 223-238. https://doi.org/10.1016/j.cvfa.2019.11.005.
- 17. Cacciò, S.M., Lalle, M., Svärd, S.G. Host specificity in the *Giardia duodenalis* species complex. Infection, genetics and evolution // Journal of molecular epidemiology and evolutionary genetics in infectious diseases. 2018. Vol. 66. P. 335-345. https://doi.org/10.1016/j.meegid.2017.12.001.
- 18. Daugschies, A., Najdrowski, M. Eimeriosis in cattle: current understanding // Journal of veterinary medicine. B, Infectious diseases and veterinary public health. 2005. Vol. 52(10). P. 417-427. https://doi.org/10.1111/j.1439-0450.2005.00894.x.
- 19. Ranasinghe, S., Armson, A., Lymbery, A.J., Zahedi, A., Ash, A. Medicinal plants as a source of antiparasitics: an overview of experimental studies // Pathogens and global health. 2023. Vol. 117(6). P. 535-553. https://doi.org/l0.1080/20477724.2023.2179454.
- 20. Assad, R., Reshi, Z.A., Mir, S.H., Rashid, I., Shouche, Y., Dhotre, D. Chapter 17 Bioprospecting appraisal of Himalayan pindrow fir for pharmacological applications, Academic Press, 2021. P. 461-482. https://doi.org/10.1016/B978-0-12-824109-7.00003-0.
- 21. Lantto, T.A., Dorman, H.J.D., Shikov, A.N., Pozharitskaya, O.N., Makarov, V.G., Tikhonov, V.P., Hiltunen, R., Raasmaja, A. Chemical composition, antioxidative activity and cell viability effects of a Siberian pine (*Pinus sibirica* Du Tour) extract // Food Chemistry. 2009. Vol. 112 (4).

- P. 936-943. https://doi.org/10.1016/j.foodchem.2008.07.008.
- 22 Artemkina, N.A., Orlova, M.A. Lukina, N.V. Chemical composition of *Juniperus sibirica* needles (*Cupressaceae*) in the forest-tundra ecotone, the Khibiny Mountains // Russ. J. Ecol. 2016. Vol. 47. P. 321-328. https://doi.org/10.1134/S106741361604007X.
- 23. Sampietro, D.A., Gomez, A.L., Jimenez, C.M., Lizarraga, E.F., Ibatayev, Z.A., Suleimen, Y.M., Catalán, C.A. Chemical composition and antifungal activity of essential oils from medicinal plants of Kazakhstan // Natural product research. 2017. -Vol. 31(12). P. 1464-1467. https://doi.org/10.1080/14786419.2016.1258560.
- 24. Caramiello, R., Bocco, A., Buffa G., Maffei M. Chemotaxonomy of *Juniperus communis*, *J. sibirica* and *J. intermedia* // Journal of Essential Oil Research. 1995. Vol. 7(2). P. 133-145. https://doi.org/10.1080/10412905.1995.9698488.
- 25. Lesjaka, M.M., Bearaa, I.N., Orčića, D.Z., Anačkovb, G.T., Baloga, K.J., Franciškovića, M.M., Mimica-Dukića, N.M. *Juniperus sibirica* Burgsdorf. as a novel source of antioxidant and anti-inflammatory agents // Food Chemistry. 2011. Vol. 124 (3). P. 850-856. https://doi.org/10.1016/j.foodchem.2010.07.006.
- 26. Zhang, Y., Wu, D., Kuang, S., Qing, M., Ma, Y., Yang, T., Wang, T., Li, D. Chemical composition, antioxidant, antibacterial and cholinesterase inhibitory activities of three Juniperus species // Natural product research. 2020. Vol. 34(24). P. 3531-3535. https://doi.org/10.1080/14786419.2019.1579811.
- 27. Farzaneh, A., Hadjiakhoondi, A., Khanavi, M., Manayi, A., Bahramsoltani, R., Kalkhorani M. *Filipendula ulmaria* (L.) Maxim. (Meadowsweet): a Review of Traditional // Research Journal of Pharmacognosy (RJP). 2022. Vol. 9(3). P. 85-106. https://doi.org/10.22127/RJP.2021.302028.1781.
- 28. <u>Čutović N., Marković T., Kostić M., roš Gašić U., Prijić Ž., Ren X., Lukić M., Bugarski B.</u> Chemical Profile and Skin-Beneficial Activities of the Petal Extracts of *Paeonia tenuifolia* L. from Serbia // Pharmaceuticals (Basel). 2022. Vol. 15(12). P. 1537. https://doi.org/10.3390/ph15121537.
- 29. Day, A.J., Bao, Y., Morgan, M.R., Williamson, G. Conjugation position of quercetin glucuronides and effect on biological activity // Free radical biology & medicine. 2000. Vol. 29(12). P. 1234-1243. https://doi.org/10.1016/s0891-5849(00)00416-0.
- 30. Lupanova I.A, Saybel O.L., Ferubko E.V., Mizina P.G. Pharmacological screening of *Serratula coronata* L. herbal extract // Problems of Biological Medical and Pharmaceutical Chemistry January. 2022. Vol. 25(3). P. 43-48. https://doi.org/10.29296/25877313-2022-03-06.
- 31. Kroma, A., Pawlaczyk, M., Feliczak-Guzik, A., Urbańska, M., Jenerowicz, D., Seraszek-Jaros, A., Kikowska, M., Gornowicz-Porowska, J. Phytoecdysteroids from *Serratula coronata L.* for Psoriatic Skincare // Molecules (Basel, Switzerland). 2022. Vol. 27(11). P. 3471. https://doi.org/10.3390/molecules27113471.
- 32. Odinokov, V.N., Galyautdinov, I.V., Nedopekin, D.V., Khalilov, L.M., Shashkov, A.S., Kachala, V.V., Dinan, L., La-

- font, R. Phytoecdysteroids from the juice of *Serratula coronata L*. (Asteraceae) // Insect biochemistry and molecular biology. 2002. Vol. 32(2). P. 161-165. https://doi.org/10.1016/s0965-1748(01)00106-0.
- 33. Vazini, H, Rahimi Esboei, B. In vitro study of the effect of hydroalcholic extracts of *Carum copticum* and *Ferula asafetida* against *Trichomonas vaginalis* // Scientific Journal of Kurdistan University of Medical Sciences. 2018. Vol. 23(1). P. 52-61. https://doi.org/10.52547/sjku.23.1.76.
- 34. Ramadan, N.I., Abdel-Aaty, H.E., Abdel-Hameed, D.M., El Deeb, H.K., Samir, N.A., Mansy, S.S., Al Khadrawy, F.M. Effect of Ferula assafoetida on experimental murine *Schistosoma mansoni* infection // Journal of the Egyptian Society of Parasitology. 2004. Vol. 34(3 Suppl). P. 1077-1094.
- 35. Tavassoli, M., Jalilzadeh-Amin, G., Fard, V. R. B., Esfandiarpour, R. The in vitro effect of *Ferula asafoetida* and *Allium sativum* extracts on *Strongylus* spp // Annals of parasitology. 2018. Vol. 64(1). P. 59-63. https://doi.org/10.17420/ap6401.133.
- 36. Arbabi, M., Haddad, A., Esmaeli, M., Hooshyar, H., Sehat, M. In Vitro Anthelmintic Effect of *Ferula assa-foe-tida* Hydroalcoholic Extract Against Flukes of *Fasciola hepatica* and *Dicrocoelium dendriticum* // Jundishapur J. Nat. Pharm. Prod. 2023. Vol. 18(1). P. e133030. https://doi.org/10.5812/jjnpp-133030.
- 37. Gundamaraju, R. Evaluation of anti-helmintic activity of *Ferula foetida* "Hing-A natural Indian spice" aqueous extract // Asian Pac. J. Trop. Dis. 2013. Vol. 3(3). P. 189-91. https://doi.org/10.1016/S2222-1808(13)60038-9.
- 38. Kirillov, V.Yu., Stikhareva, T.N., Serafimovich, M.V., Mukasheva, F.T., Gering, A.V., Sarsenbaeva, L.A., Atazhanova, G.A., Adekenov, S.M. Chemical composition of essential oil from two species of *Pulsatilla* growing wild in Northern Kazakhstan // Bulletin of the Karaganda university. 2018. Vol. 2(90). P. 29-34.
- 39. Bajtenov M.S. Flora Kazahstana. Illjustrirovannyj opredelitel' semejstv i rodov. (Flora of Kazakhstan. Illustrated guide to families and genera) Almaty: Almaty kitap, 1978. 305 s.
- 40. The Plant List. Version 1.1. Electronic resource: http://www.theplantlist.org.
- 41. State Pharmacopeia of Kazakhstan Republic. Almaty, 2008. Vol. 1. P. 361-401.
- 42. Aidarkhanova, G.S. Research report (AP05136154), Nur-Sultan, 2021. 109 pp. (in Russian).
- 43. Nikolić, B., Vasiljević, B., Mitić-Ćulafić, D., Lesjak, M., Vuković-Gačić, B., Mimica Dukić, N., Knežević-Vukčević, J. Screening of the antibacterial effect of *Juniperus sibirica* and *Juniperus sabina* essential oils in a microtitre platebased MIC assay // Botanica Serbica. 2016. Vol. 40(1). P. 43-48. https://doi.org/10.5281/zenodo.48858.
- 44. Elaine M Aldred BSc (Hons), DC, Lic Ac, Dip Herb Med, Dip CHM. Pharmacology: A Handbook for Complementary Healthcare Professionals, 2009. https://doi.org/10.1016/B978-0-443-06898-0.X0001-1.
- 45. Zahra, N., Iqbal, J., Arif, M., Abbasi, B.A., Sher, H., Nawaz, A.F., Yaseen, T., Ydyrys, A., Sharifi-Rad, J., Ca-

lina, D. A comprehensive review on traditional uses, phytochemistry and pharmacological properties of Paeonia emodi Wall. ex Royle: current landscape and future perspectives // Chinese medicine. – 2023. – Vol. 18(1). – P. 23. https://doi.org/10.1186/s13020-023-00727-7.

46. Akhmetkarimova, Z., Temirgaziev, B. Phytoecdysteroids: Chemical Structure And Biological Activity // Eurasian Journal of Applied Biotechnology. – 2021. – Vol. 2. – P.

3-22. https://doi.org/10.11134/btp.2.2021.1.

47. Aidarkhanova, G., Kubentayev, S.A., <u>Kukhar, Ye.V.</u>, Urazalina, A.S. Current state of populations, resources, and biological activity *Pulsatilla patens* (Ranunculaceae) in Northern Kazakhstan // Biodiversitas. – 2022. – Vol. 23 (5). – P. 2311-2320. https://doi.org/10.13057/biodiv/d230509.

УДК 616:619. 615.322-615.281.9:615.284-615.282.84-615.281.9

АНАЛИЗ АНТИПАРАЗИТАРНОЙ АКТИВНОСТИ ДИКОРАСТУЩИХ РАСТЕНИЙ ВОСТОЧНОГО И СЕВЕРНОГО КАЗАХСТАНА

Кухар Е.В.^{1*}, Айдарханова Г.С.²

- ¹ Научно-исследовательская платформа сельскохозяйственной биотехнологии, Казахский агротехнический исследовательский университет им. С. Сейфуллина, Казахстан, г. Астана
- ² Кафедра биология, защита и карантин растений, Казахский агротехнический исследовательский университет им. С. Сейфуллина, Казахстан, г. Астана
- *Автор-корреспондент, Кухар Е.В., kucharev@mail.ru.

АБСТРАКТ

Различные гельминтозы и иные паразитарные инфекции широко распространены среди поголовья сельскохозяйственных животных. Гельминтозная инфекция обычно лечится антигельминтными препаратами, но резистентность к антигельминтным препаратам становится все более серьезной проблемой. Изучение потенциальных растительных противопаразитарных средств, которые недороги и широко распространены, может быть многообещающей альтернативой. Установлено наличие антигельминтных свойств у масляных, водно-спиртовых и большинства водных экстрактов Filipendula vulgaris Moench., Pinus sylvestris, Picea abies L., Ábies sibírica, Pinus sibirica DuTour, Juniperus communis L., Paeonia anomala L. с лесных экосистем Восточного Казахстана; Ferula tatarica Fisch., Pulsatilla uralensis (Zămels) Tzvel., (Serratula coronata L. из экосистем Северного Казахстана. Наибольшее влияние и быструю гибель аннелид вызывали спиртовые настойки, наименьшее – водные экстракты. Лучшими антигельминтными свойствами отличались водно-спиртовые экстракты подземной части Прострела желтеющего (Pulsatilla uralensis (Zămels) Tzvel.), спиртовая настойка надземной части Серпухи венценосной (Serratula coronata L.). Впервые описана противопаразитарная активность надземной и подземной части Прострела желтеющего (Pulsatilla uralensis (Zămels) Tzvel.), произрастащего в экосистемах Северного Казахстана.

Ключевые слова: антипаразитарная активность, антигельминтные свойства, хвойные деревья, кустарники, травянистые растения

ӘОЖ 616:619. 615.322-615.281.9:615.284-615.282.84-615.281.9

ШЫҒЫС ЖӘНЕ СОЛТҮСТІК ҚАЗАҚСТАНДАҒЫ ЖАБАЙЫ ӨСІМДІКТЕРДІҢ ПАРАЗИТТЕРГЕ ҚАРСЫ БЕЛСЕНДІЛІГІН ТАЛДАУ

Кухар Е.В.^{1*}, Айдарханова Г.С.²

- ¹ Ауылшаруашылық биотехнологиясы бойынша ғылыми-зерттеу платформасы, С.Сейфуллин атындағы Қазақ агротехникалық зерттеу университеті, Қазақстан, Астана қ.
- ² С.Сейфуллин атындағы Қазақ агротехникалық зерттеу университетінің биология, өсімдіктерді қорғау және карантин кафедрасы, Қазақстан, Астана қ.
- *Автор-корреспондент, Кухар Е.В., kucharev@mail.ru.

АНДАТПА

Ауыл шаруашылығы жануарлары арасында әртүрлі гельминтоздар және басқа паразиттік инфекциялар кең таралған. Жүрек құртының инфекциясы әдетте антигельминтикалық препараттармен емделеді, бірақ антигельминтикалық препараттарға төзімділік барған сайын күрделі мәселе болып табылады. Қымбат емес және кеңінен қол жетімді

әлеуетті шөпке қарсы антипаразиттік агенттерді зерттеу перспективалы балама болуы мүмкін. Антельминтикалық қасиеттердің болуы Filipendula vulgaris Moench., Pinus sylvestris, Picea abies L., Ábies sibírica, Pinus sibirica DuTour, Juniperus communis L., Paeonia anomala L. e. Шығыс Қазақстан; Ferula tatarica Fisch., Pulsatilla uralensis (Zămels) Tzvel., (Serratula coronata L. Солтүстік Қазақстан экожүйелерінен. Анелидтердің ең үлкен әсері және тез өлуі алкоголь тұнбаларымен, ең азы - сулы сығындылармен болды. Ең жақсы антигельминтикалық қасиеттері сарғыш лумбагоның (Pulsatilla uralensis (Zămels) Tzvel.) жер асты бөлігінің сулы-спирттік сығындыларымен және тәжді серпуханың (Serratula coronata L.) әуе бөлігінің спиртті тұнбаларымен ерекшеленді. Алғаш рет Солтүстік Қазақстанның экожүйесінде өсетін сарғыш любагоның (Pulsatilla uralensis (Zămels) Tzvel.) жер үсті және жер асты бөліктерінің паразитке қарсы әрекеті сипатталды.

Түйін сөздер: паразиттерге қарсы белсенділігі, антигельминтикалық қасиеттері, қылқан жапырақты ағаштар, бұталар, шөптесін өсімдіктер.